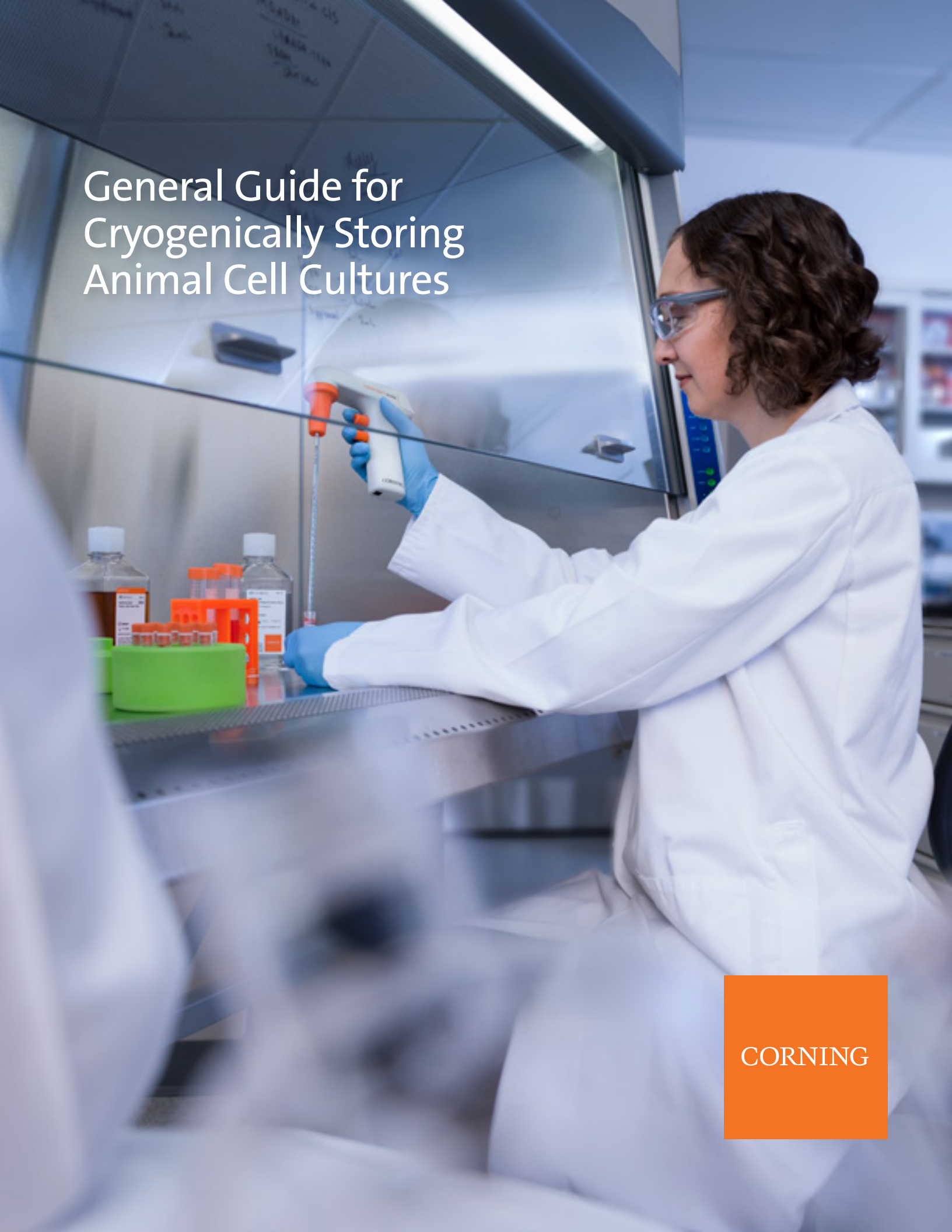


General Guide for Cryogenically Storing Animal Cell Cultures



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Introduction

Maintaining healthy, growing cell cultures is a demanding task made more difficult by the everpresent risk of their loss through accidents or contamination. In addition, actively growing cell cultures are not static but, like all populations of microorganisms, subject to age-related or environmentally-induced changes which can result in their ongoing evolution and potential loss.

These problems can be reduced by using cryogenic preservation to stop biological time for cell cultures, effectively putting them into true suspended animation. This concept, long a favorite ploy of science fiction writers and movie producers, has been a reality since the important discovery by Polge, Smith and Parkes¹⁴ in 1949 that glycerol prevents injury to cells caused by freezing. Many cook book-style protocols are available for freezing cells and these procedures usually perform well^{4,7,17-20}. Recently, specialized protocols for freezing cell pellets, tissues, hematopoietic stem cells, and mesenchymal stem cells have also been developed^{3,9,12,15}. It is essential, however, when problems arise or protocol adaptations and improvements must be made, that the underlying concepts on which they are based are well understood. This guide examines both the basic theoretical concepts and practical aspects necessary for successfully freezing animal cells and managing a cell repository.

Advantages of Freezing Cell Cultures

Once successfully frozen and stored, cell cultures require little time and effort for their maintenance. The only real cost is the expense of maintaining an ultracold (-130°C or lower) mechanical freezer or liquid nitrogen supply. This limited expense compares very favorably with the time, effort, and substantial cost of the media and supplies necessary for maintaining actively growing cultures, or for the cost of obtaining a new culture from a repository. Frozen cultures provide an important backup supply for replenishing occasional losses due to contamination or accidents and provide the assurance of a homogeneous culture supply. Cellular changes or alterations occur in all actively growing populations. These changes often result in the loss of important characteristics during evolution of the cultures thereby introducing unwanted variables into long-term experiments. Cryogenically preserved cultures apparently do not undergo any detectable changes once they are stored below -130°C^{1,10}. Therefore, the biological effects of *in vitro* cellular aging and evolution may be minimized by frequently returning to frozen stock cultures, allowing ongoing long-term culture experiments to be successfully completed without these unwanted variables. Frozen cultures also provide a valuable baseline against which future experimentally-induced changes may be compared or measured.

General Events During Cell Freezing

To understand why freezing protocols work, it is necessary to examine both the intracellular and extracellular events occurring in animal cell cultures during the freezing process^{2,5,10}. Initial cooling from room temperature to 0°C slows cellular metabolism, rapidly disrupting active transport and ionic pumping. This disruption usually does not result in cellular damage if the culture medium is osmotically balanced. As cooling continues (0°C to -20°C) ice crystals begin to form in the extracellular environment which increases the solute concentration of the culture medium. As a result, water begins to move out of the cells and into the partially frozen extracellular medium, beginning the process of cellular dehydration and shrinkage.

When the cooling process is rapid, intracellular ice crystals form before complete cellular dehydration has occurred. These ice crystals disrupt cellular organelles and membranes and lead to cell death during the recovery (thawing) process (Figure 1).

When the cooling process is slow, free intracellular water is osmotically pulled from the cells resulting in complete cellular dehydration and shrinkage. This can also cause cellular death, but there is little agreement on the mechanisms involved. The physical stresses of cellular shrinking may cause some damage resulting in irreparable membrane loss and cytoskeletal and organelle disruption. Damage may also be caused by the high concentrations of solutes in the remaining unfrozen extracellular medium (essentially a brine solution). These solutes attack cells both externally and internally, resulting in membrane damage, pH shifts, and general protein denaturation.

Advantages of Freezing Cultures

- ▶ Serves as a backup supply for emergencies.
- ▶ Reduces risk of microbial contamination and cross-contamination with other cell lines and genetic drift changes.
- ▶ Provides a more homogeneous population by minimizing culture aging and evolution.

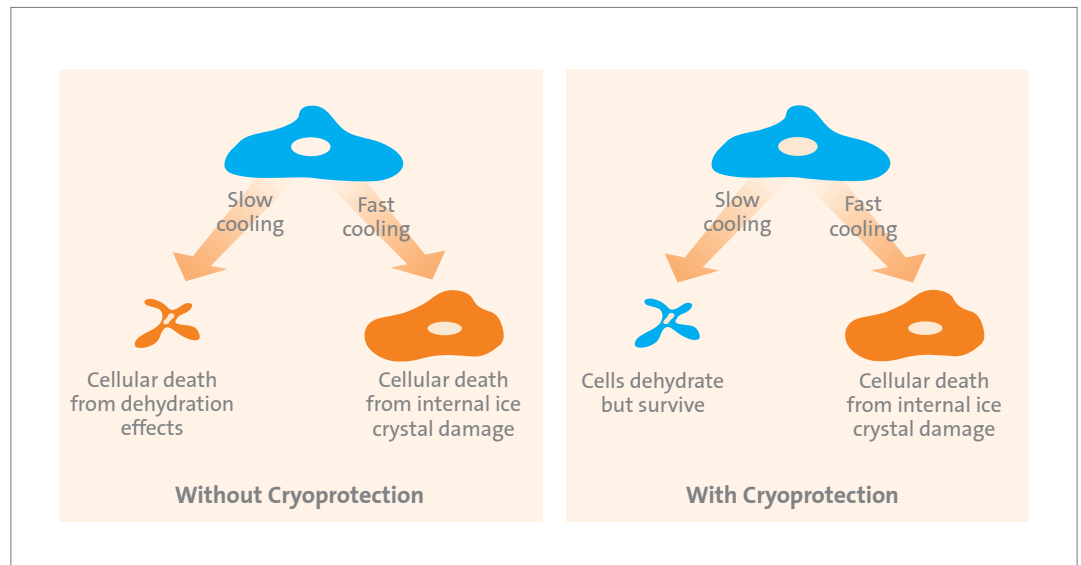


Figure 1. Effects of freezing rates on cells.

However, when the cooling rate is slow enough to prevent intracellular ice formation, but fast enough to avoid serious dehydration effects, cells may be able to survive the freezing and thawing process. Since this can be difficult to control manually, it is recommended to use a device to control the freezing rate, such as a Corning® CoolCell container. This survival zone or window is readily observed in many bacteria and other prokaryotes, but for most eukaryotic cells it is nonexistent or very difficult to find without using cryoprotective agents. These agents have little effect on the damage caused by fast freezing (intracellular ice crystal formation), but rather prevent or lessen the damage caused by slow freezing (dehydration and shrinkage)¹⁰.

The final storage temperature is also critical for successful cryopreservation. To completely stop biological time, storage temperatures must be maintained below -130°C , the glass transition point below which liquid water does not exist and diffusion is insignificant. While many cell cultures are successfully stored at -70°C to -90°C for months or even years, biological time is not stopped, only slowed, and cellular damage or changes will accumulate.

Storage in liquid nitrogen at -196°C effectively prevents all thermally driven chemical reactions. Only photo-physical effects caused by background ionizing radiation still operate at this temperature. Thousands of years are estimated to be necessary before background radiation will have a noticeable effect on cryopreserved cultures^{2,10}.

Practical Aspects of Cell Freezing

Under the best of circumstances the process of freezing remains stressful to all cell cultures. It is important that everything possible be done to minimize these stresses on the cultures in order to maximize their subsequent recovery and survival. The following suggestions and recommendations are designed to augment the protocols referred to earlier.

Cell Selection

First ensure the cells are in their best possible condition. Select cultures near the end of log phase growth (approximately 90% confluent), and change their medium 24 hours prior to harvesting. Carefully examine the culture for signs of microbial contamination. Facilitate this by growing cultures in antibiotic-free medium for several passages prior to testing. This allows time for any hidden, resistant contaminants (present in very low numbers) to reach a higher, more easily detected level. Samples of these cultures are then examined microscopically and tested by direct culture for the presence of bacteria, yeasts, fungi, and mycoplasmas.

Mycoplasmas present a special problem since they can be found in cultures at very high concentrations (up to 10^8 organisms per milliliter of medium) without any visible effects or turbidity. As a result, as many as 20% of all animal cell cultures are contaminated by these ubiquitous but unseen organisms. Although special efforts are required to detect mycoplasmas, the serious consequences of their presence makes testing frozen culture stocks absolutely essential^{11,16}.

Check for both the identity of the cultures and the presence of any expected special characteristics. Monitor cell identities by karyology and isoenzyme analysis, ensuring that they are, at the very least, the correct species¹³.

Cell Harvesting

Start with the standard harvesting procedure generally recommended for the culture, and be as gentle as possible. Remove all dissociating agents by washing or inactivation (especially important when using serum-free medium). Centrifugation, when absolutely necessary, should only be hard enough to obtain a soft pellet; 100 x g for 5 to 6 minutes is usually sufficient. To ensure uniformity of the final frozen stock, pool the contents of all harvested culture vessels. This also makes it easier to perform essential quality control testing for microbial contamination and culture identity.

Count and then dilute or concentrate the harvested cell suspension to twice the desired final concentration, which is usually 4 to 10 million viable cells per milliliter. An equal volume of medium containing the cryoprotective agent at twice its final concentration will be added later to achieve the desired inoculum. Keep the cells chilled to slow their metabolism and prevent cell clumping. Avoid alkaline pH shifts by gassing with CO₂ when necessary.

Prevent Freezing Damage

- ▶ Use slow freezing to remove all intracellular water.
- ▶ Use cryoprotective agents to minimize dehydration effects.
- ▶ Store below -130°C to completely stop biological time.

Cryoprotection

As mentioned earlier, cryoprotective agents are necessary to minimize or prevent the damage associated with slow freezing. The mechanisms providing this protection, although not completely understood, appear to work primarily by altering the physical conditions of both the ice and solutions immediately surrounding (external to) the cells. Permeation of the cells by cryoprotectants does not appear to be necessary for their proper functions⁵. Remember, protection against fast freezing damage (internal ice formation) is not provided by these agents, but rather by careful control of the freezing rate. A wide variety of chemicals provide adequate cryoprotection, including methyl acetamide, methyl alcohol, ethylene glycol and polyvinyl pyrrolidone⁸. However, dimethylsulfoxide (DMSO) and glycerol are the most convenient and widely used. Many of these agents, although providing excellent cryoprotection, have toxic side effects on cultures making their use difficult.

DMSO is most often used at a final concentration of 5% to 15% (v/v). Always use reagent or other high purity grades that have been tested for suitability. Sterilize by filtration through a 0.2 micron nylon membrane in a polypropylene or stainless steel housing, and store in small quantities (5 mL). **CAUTION:** Take special care to avoid contact with solutions containing DMSO. It is a very powerful polar solvent capable of rapidly penetrating intact skin and carrying in with it harmful contaminants such as carcinogens or toxins. Some cell lines are adversely affected by prolonged contact with DMSO. This can be reduced or eliminated by adding the DMSO to the cell suspension at 4°C and removing it immediately upon thawing. If this does not help, lower the concentration or try glycerol or another cryoprotectant.

Glycerol is generally used at a final concentration of between 5% and 20% (v/v). Sterilize by autoclaving for 15 minutes in small volumes (5 mL) and refrigerate in the dark. Although less toxic to cells than DMSO, glycerol frequently causes osmotic problems, especially after thawing. Always add it at room temperature or above and remove slowly by dilution.

High serum concentrations may also help cells survive freezing. Replacing standard media-cryoprotectant mixtures with 95% serum and 5% DMSO may be superior for some overly sensitive cell lines, especially hybridomas.

Add cryoprotective agents to culture medium (without cells) immediately prior to use to obtain twice the desired final concentration (2X). Mix this 2X solution with an equal volume of the harvested cell suspension (also 2X) to obtain the inoculum for freezing. This method is less stressful for cells, especially when using DMSO as the cryoprotectant.

Storage Vessels

After the cryoprotective solution is mixed with the cell suspension, the resulting inoculum is added in small aliquots (usually 1 to 2 mL) to each storage vessel. Due to the extremely low temperatures encountered during cryogenic storage, not all vessel materials or designs are suitable or safe. Many materials become very brittle at these temperatures; vessels made from them may shatter or crack during storage or thawing. Carefully check the vessel manufacturers' recommendations on proper selection and use.

Also important is selecting the sealing system or cap design used to maintain the integrity of the vessel, especially for storage in liquid nitrogen. If these vessels leak during storage (as many do*), they will slowly fill with liquid nitrogen. When they are eventually returned to room temperature, the liquid nitrogen quickly vaporizes causing a rapid pressure buildup. The vessels may then violently blow off their caps or explode to vent the pressure and release their contents into the atmosphere. This is a very dangerous situation, especially if the vessels contained pathogenic organisms or potentially toxic or harmful substances.

WARNING: To avoid injury, DO NOT immerse plastic or glass cryogenic vials in liquefied gases such as liquid nitrogen. Vials immersed in liquefied gases can develop leaks. When such vials return to room temperature, rapid pressure can build up and shatter the vials, or cap seals. Harmful or biohazard materials contained in the vials can release during the venting of pressurized gas. Always store vials only in the vapor phase ABOVE any liquefied gas.



Figure 2. Corning externally threaded (left) and internally threaded (right) cryogenic vials.

Two types of vessels are commonly used for cryogenic storage: heat-sealable glass ampules and plastic (usually polypropylene) screw-capped vials. Both are available in a variety of sizes (1 to 5 mL capacity) although the smaller sizes are preferred for cryogenic storage (see Figure 2).

Because of sealing and labeling problems, glass ampules are no longer widely used in cell culture laboratories. Invisible pinhole leaks may occur in vials during the sealing process; if these are later stored submerged in liquid nitrogen, they may explode when removed for thawing. Pinholes can usually be detected before freezing by immersing sealed ampules for 30 minutes in a chilled solution of 70% ethanol containing 1% methylene blue. This solution will rapidly penetrate and stain any leaky ampules; after rinsing with water, defective ampules are then easily detected and discarded.

Due to their greater safety and convenience, plastic vials have largely replaced glass ampules for cryogenic storage. The wide variety of styles and special features such as printed marking areas and colored caps for easier identification also add to their popularity.

Several cap styles are available, some with an internally threaded cap, and others with externally threaded designs which help minimize contamination (see Figure 3). Cryogenic bags are also available for larger volumes and are unique containers that remain flexible at ultra-low temperatures.

Labeling and Recordkeeping

Providing for long-term location and identification of frozen cultures is the most frequently overlooked area of cryogenic storage. A cryogenic cell repository is expected to outlast the laboratory workers who contribute to it, but poorly maintained or missing inventory records, and improperly or illegibly labeled vials and ampules may prevent this, especially after the people responsible have gone.

Labels must contain enough information to locate the appropriate records; usually the culture's identity, date frozen, and initials of the person responsible are sufficient. Most plastic vials have printed marking spots or areas for easy labeling. On vials and ampules without marking spots, use cloth labels with special adhesives formulated for cryogenic conditions.

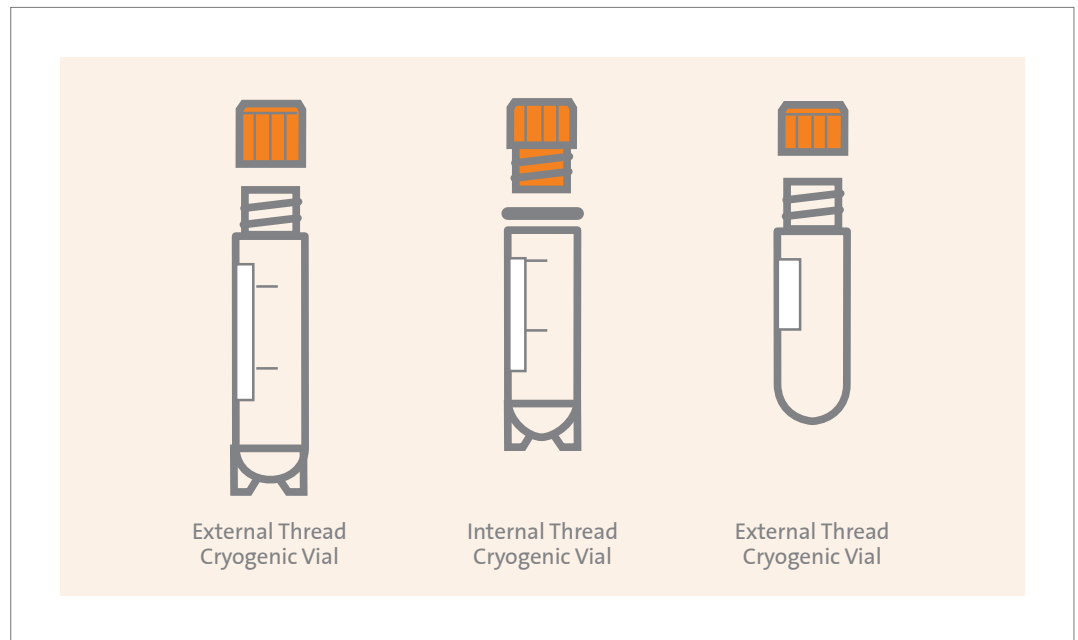


Figure 3. Corning cryogenic vial cap designs

Special ceramic-based inks are available for labeling glass ampules. These are applied prior to filling and then baked onto the glass, usually during dry heat sterilization. Permanent marking spots can be applied on glass ampules with white nail polish. A laboratory marking pen is then used to write on the spot once it has dried.

No matter which labeling method is chosen, use special care to check its permanency under cryogenic conditions. Some marking spots, inks, and labels may flake off or fade during long-term storage; a trial run of at least several weeks is recommended.

Fully detail in the records the culture's storage conditions, including all of the following information: culture identity, passage or population doubling level, date frozen, freezing medium and method used, number of cells per vial, total number of vials initially frozen, and the number remaining, their locations, their expected viability, and results of all quality control tests performed (sterility, mycoplasma, species, karyotype, etc.). Additional culture information, especially their origin, history, growth parameters, special characteristics, and applications, is also helpful and should be included whenever possible.

Make special efforts to keep all records up-to-date and to ensure everyone in the facility is properly using them. Use preprinted forms to make the information recording process easier and more likely to be completed. Keep updated, duplicate copies of all critical records in a safe place removed from the laboratory area to guard against their accidental loss or destruction. This is especially important if a computer-based recordkeeping system is used; a current back up copy should always be maintained in addition to the information stored in the computer.

Cooling Rate

The cooling rate used to freeze cultures must be just slow enough to allow the cells time to dehydrate, but fast enough to prevent excessive dehydration damage. A cooling rate of -1°C to -3°C per minute is satisfactory for most animal cell cultures. Larger cells, or cells having less permeable membranes may require a slower freezing rate since their dehydration will take longer.

The best way to control cooling rates is using electronic programmable freezing units. Although expensive, they allow precise control of the freezing process, give very uniform and reproducible results, and can freeze large numbers of vials or ampules. Most units are available with chart recorders for a permanent record of the cooling process.

There are a variety of mechanical freezing units that provide adequate control of the cooling rate and are relatively inexpensive. Some units use racks designed to hold vials at predetermined depths in the neck of a liquid nitrogen freezer. The cooling rate is dependent on the total number of vials and the depth at which the rack is placed. Another design uses an alcohol-filled metal or plastic canister containing a rack with a capacity of up to 24 vials. The filled canister is placed in an ultracold mechanical freezer where the alcohol acts as a bath to achieve more uniform heat transfer and cooling. After freezing 4 to 5 hours, the vials are removed from the canister and transferred to their final storage locations.

A Corning® CoolCell® foam container, in combination with a -80°C freezer, will provide alcohol-free freezing at the rate of -1°C/minute that is ideal for cryopreservation of most cells and cell lines. Using a combination of uniform-density cross-linked polyethylene foam, a solid state core, and radial vial symmetry, freezing profiles are consistent and reproducible. The foam is non-absorbent and will impose negligible change in the freezer environment; thereby protecting nearby frozen samples. The removal of vials is accomplished by tipping the device upside down so the vials slide out. These devices are easy to use, are alcohol- and fluid-free, reproducible, and offer a simple consistent way to standardize controlled-rate freezing.

Insulated cardboard or polystyrene foam boxes are commonly used as freezing chambers in ultracold freezers. These homemade devices work well with many cell lines but do not always give controlled, reproducible, or uniform cooling. As a result, there may be serious differences in viability among the vials upon thawing. This homemade approach is not recommended for valuable or irreplaceable cultures.

No matter which cooling method is used, transfer from the cooling chamber or device to the final storage location must be done quickly to avoid warming of the vials. Use an insulated container filled with dry ice or liquid nitrogen as a transfer vessel to ensure that the cells remain below -70°C.

Cryogenic Storage

Only freezers capable of continually maintaining temperature below -130°C should be considered for long-term cryogenic storage. Although most liquid nitrogen-cooled freezers and some specially designed mechanical freezers meet this requirement, most cell culture laboratories prefer liquid nitrogen freezers. The final choice is often based on the availability of a reliable supply of liquid nitrogen, the storage capacity required, and the size of the budget. Liquid nitrogen freezers permit storage either in the vapor phase above the liquid at a temperature between -140°C and -180°C, or submerged in the liquid at a temperature below -196°C. Using vapor phase storage greatly reduces the possibility of leaky vials or ampules exploding during removal. However, since the amount of liquid nitrogen in the freezer is reduced to provide space for vapor phase storage, the freezer's holding time (the period it can maintain its storage temperature without adding more liquid nitrogen) is also reduced. This lowers the freezer's margin of safety and requires more frequent monitoring and filling. Give careful consideration to these safety issues when deciding upon a storage method.

WARNING: To avoid injury, DO NOT immerse plastic or glass cryogenic vials in liquefied gases such as liquid nitrogen. Vials immersed in liquefied gases can develop leaks. When such vials return to room temperature, rapid pressure can build up and shatter the vials, or cap seals. Harmful or biohazard materials contained in the vials can release during the venting of pressurized gas. Always store vials only in the vapor phase ABOVE any liquefied gas.

Frequently check nitrogen levels in freezers; a schedule should be established and strictly adhered to. Nitrogen evaporation is dependent on both the degree of use and the static holding time of the freezer. Sudden, unexplained increases in the evaporation rate may signal damage to the insulation or other problems with the freezer and must be carefully investigated. Avoid frost or ice buildup around freezer openings; this increases the nitrogen evaporation rate and can cause elevated temperatures in the upper portion of vapor phase freezers. Audible alarm systems for detecting low liquid nitrogen levels are available to provide additional safeguards; however, they provide a false sense of security if not monitored 24 hours a day.

Thawing

CAUTION: Always use appropriate safety equipment when removing vials and ampules from liquid or vapor phase nitrogen freezers. A full face shield, heavy gloves, and lab coat are strongly recommended for protection against exploding vials or ampules.

Remove the vial or ampule from its storage location and carefully check both the label and storage record to ensure that it is the correct culture. Place the vessel in warm water, agitating gently until completely thawed. Rapid thawing (60 to 90 seconds at 37°C) provides the best recovery for most cell cultures; it reduces or prevents the formation of damaging ice crystals within cells during rehydration.

Recovery

Since some cryoprotective agents may damage cells upon prolonged exposure, remove the agents as quickly and gently as possible. Several approaches are used depending on both the cryoprotective agents and characteristics of the cells. Most cells recover normally if they have the cryoprotective agent removed by a medium change within 6 to 8 hours of thawing. Transfer the contents of the ampule or vial to a 75 cm² flask or other suitable vessel containing 0.2-0.3 milliliters per cm² of culture medium and incubate normally. As soon as a majority of the cells have attached, remove the medium containing the now diluted cryoprotective agent and replace with fresh medium.

For cells that are sensitive to cryoprotective agents, removing the old medium is easily accomplished by gentle centrifugation. Transfer the contents of the vial or ampule to a 15 mL centrifuge tube containing 10 mL of fresh medium and spin for 5 minutes at 100 x g. Discard the supernatant containing the cryoprotectant and resuspend the cell pellet in fresh medium. Then transfer the cell suspension to a suitable culture vessel and incubate normally.

When glycerol is used as the cryoprotectant, the sudden addition of a large volume of fresh medium to the thawed cell suspension can cause osmotic shock, damaging or destroying the cells. Use several stepwise dilutions with an equal volume of warm medium every 10 minutes before further processing to give the cells time to readjust their osmotic equilibrium.

Problem Solving Suggestions

Viability problems associated with cryogenic storage are usually noticed soon after cultures are thawed and plated. There are four major areas where problems occur:

During harvesting and processing of the cells

Problems may be caused by excessive exposure of the cells to dissociating agents; using a cryoprotective agent that is toxic; or allowing high density cell suspensions to remain too long at room temperature or at a pH that is too basic.

During the cooling (freezing) process

Excessive cell damage and reduced culture viability often result from using a cooling rate that is too fast or too slow, or when the cooling process is temporarily interrupted. Not using a suitable cryoprotective agent at an appropriate concentration will also result in viability problems.

During cyrogenic storage

Culture viability is often reduced when vials are allowed to warm up during transfer to the freezer, or if the repository temperature is not consistently maintained at appropriate cryogenic temperatures.

During thawing and recovery

Problems arise when the thawing process is too slow or the cryoprotectants are improperly removed (see above).

These viability problems can often be corrected by using the following technique to identify the stage in the freezing process where the problem originates.

Harvest enough cells to prepare at least four vials. Then remove a sample of cell suspension, equivalent in cell number to that which will be placed into the vials, and immediately place it into a culture vessel with an appropriate amount of medium and incubate. This culture will be used as a control to compare with the cultures set up in the remaining steps.

Freezing Checklist

- ▶ Gently harvest cultures.
- ▶ Check cultures for contamination, especially mycoplasma.
- ▶ Check culture identity by karyotyping or isoenzyme analysis.
- ▶ Use tested cryoprotective agents.
- ▶ Only use vials tested for cryogenic conditions.
- ▶ Ensure labels are permanent and complete.
- ▶ Control the cooling rate.
- ▶ Store cultures below -130°C.
- ▶ Monitor liquid nitrogen levels frequently.
- ▶ Keep good records.

Next, add the cryoprotective agent to the remaining cells and divide among three vials. Place one vial at 4°C for one hour. Then remove the cells from the vial, process as though they had just been thawed from the freezer, and plate in medium as above. This culture will be compared with the control culture to determine if there are any problems associated with cryoprotective agent.

Meanwhile, process the remaining vials through the slow cooling process as usual. One vial is then immediately thawed and processed as above. This culture will be compared with the control culture to determine if there are any problems associated with the slow cooling process.

The remaining vial is then transferred to the cryogenic freezer and stored overnight before being thawed and processed as above. This culture will be compared with the control culture to determine if there are any problems associated with the cryogenic storage conditions. If additional vials of cells are available, several different recovery techniques should be used to determine if the recovery technique is the source of the problem.

By comparing all of the cultures to the original culture, it should then be possible to determine at which stage of the freezing process the problem occurred. Once this is known, the information presented in this guide and its references should be enough to eliminate the problem.

Managing a Cell Repository

The effort and expenses of managing a repository should be kept in line with the value of the cultures stored within it. This value is determined by answering two questions: How much time, money, and effort is already invested in these stored cell cultures? And, what are the consequences of losing them? Cultures that are easily replaced through other labs or commercial sources may not require special efforts, but unique cultures, such as hybridomas and other genetically engineered cells, are irreplaceable and require that special efforts be made to ensure their safety. The answers to these questions will help determine just how extensive and thorough your efforts should be.

Next, identify the potential problem areas that can cause the loss of these cultures. Some of these areas, such as vessel selection, recordkeeping, labeling, freezer monitoring, storage conditions, and quality issues (contamination and species identity), have already been discussed in this guide. Decide what steps are necessary to eliminate or minimize these problems. Split irreplaceable or extremely valuable cultures among several freezers, with at least one freezer in a separate location to protect against fire or other natural disasters. Colleagues in other labs or buildings may be able to provide good backup storage, especially if a reciprocal arrangement is made for them.

One final step remains; plan ahead for emergencies! One of the most serious and unexpected emergencies is the failure of a cryogenic freezer. Careful monitoring of the liquid nitrogen level or charting the temperature may give an early warning that failure is occurring, but middle of the night failures can and do happen. Have plans prepared in advance to deal with freezer failure and other problems. If these involve a colleague's equipment, get permission and make all necessary arrangements in advance — late night phone calls are usually not appreciated.

This information has been compiled to provide a guide for better understanding of the cryogenic preservation process. For additional assistance in this area, visit www.corning.com/lifesciences, or contact Corning Scientific Support at 1.800.492.1110; outside the United States, call +1.978.442.2200; or email ScientificSupport@corning.com.

Acknowledgment

Corning wishes to acknowledge the contributions of John Ryan to this article.

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Ordering Information

Corning® Cell Culture Flasks

Cat. No.	Surface Area (cm ²)	Flask Style	Neck Style	Cap Style	Qty/Pk	Qty/Cs
430639	25	Rectangular	Canted	Vent	20	200
430641U	75	U-shaped	Canted	Vent	5	100
430825	150	U-shaped	Canted	Vent	5	50
431080	175	U-shaped	Canted	Vent	5	50
431082	225	Rectangular	Canted	Vent	5	25
10020	1,720	Corning HYPERFlask®	-	-	4	4
10030	1,720	Corning HYPERFlask®	-	-	1	4

Costar® Multiple Well Plates, Tissue Culture-treated

Cat. No.	Description	Plate Type	Qty/Pk	Qty/Cs
3516	6-well	Clear	1	50
3513	12-well	Clear	1	50
3526	24-well	Clear	1	50
3548	48-well	Clear	1	100
3596	96-well	Clear	1	50
3595	96-well	Clear*	1	50
3599	96-well	Clear	1	100
3916	96-well	Black	20	100
3917	96-well	White	20	100
3603	96-well	Black/Clear-bottom	1	48
3610	96-well	White/Clear-bottom	1	48

*Low evaporation lid

Corning Dishes, Tissue Culture-treated

Cat. No.	Dish Style* (mm)	Approx. Height (mm)	Growth Area (cm ²)	Qty/Pk	Qty/Cs
430165	35	10	8	20	500
430166	60	15	21	20	500
430167	100	20	55	20	500
430599	150	25	148	5	60
431110	245	25	500	4	16

*Dish Style (mm) 35 mm = 34.4 mm; 60 mm = 52.1 mm; 100 mm = 83.9 mm; 150 mm = 139.1 mm. Square dishes have interior bottom dimensions 224 x 224 mm.

Corning Cell Scrapers

Cat. No.	Description	Blade Length (cm)	Handle Length (cm)	Qty/Pk	Qty/Cs
3008	Cell lifter	1.9	18	1	100
3010	Scraper, small	1.8	25	1	100
3011	Scraper, large	3	39	1	100

Corning Cell Strainers

Cat. No.	Description	Qty/Pk	Qty/Cs
431750	40 µm, blue	1	50
431751	70 µm, white	1	50
431752	100 µm, yellow	1	50

Corning Cryogenic Vials

Cat. No.	Capacity (mL)	Style	Self-standing	Qty/Pk	Qty/Cs
430659	2	Round bottom, internal thread	Yes	50	500
430488	2	Round bottom, external thread	Yes	50	500

Falcon® Cell Culture Flasks, Tissue Culture-treated

Cat. No.	Surface Area (cm ²)	Flask Style	Neck Style	Cap Style	Qty/Pk	Qty/Cs
353018	25	Rectangular	Canted	Vent	20	100
353136	75	Rectangular	Canted	Vent	5	60
355001	150	Rectangular	Canted	Vent	5	40
353112	175	Rectangular	Straight	Vent	5	40
353138	225	Rectangular	Canted	Vent	5	30
353143	525	3-Layer	-	Vent	2	12
353144	875	5-Layer	-	Vent	1	8

Falcon Cell Culture Plates, Tissue Culture-treated

Cat. No.	Description	Plate Type	Qty/Pk	Qty/Cs
353046	6-well	Clear	1	50
353043	12-well	Clear	1	50
353047	24-well	Clear	1	50
353078	48-well	Clear	1	50
353072	96-well	Clear	1	50

Falcon Cell Culture Dishes

Cat. No.	Dish Style (mm)	Approx. Height (mm)	Growth Area (cm ²)	Qty/Pk	Qty/Cs
353001	35	10	11.78	20	500
353002	60	15	21.29	20	500
353003	100	20	58.95	20	200
353025	150	25	156.36	10	100

Falcon Cell Strainers

Cat. No.	Description	Qty/Pk	Qty/Cs
352340	40 µm, blue	1	50
352350	70 µm, white	1	50
352360	100 µm, yellow	1	50

Falcon Cell Scrapers

Cat. No.	Description	Qty/Pk	Qty/Cs
353085	1.8 cm TPE blade with 18 cm polystyrene handle	1	100
353086	1.8 cm TPE blade with 25 cm polystyrene handle	1	100
353089	3.0 cm TPE blade with 25 cm polystyrene handle	1	100
353087	3.0 cm TPE blade with 40 cm polystyrene handle	1	100

Corning® CoolCell® Containers

Cat. No.	Description	Capacity (Vials)	Exposed Vial Tops	Qty/Pk	Qty/Cs
432000	CoolCell, purple	12	No	1	1
432001	CoolCell LX, purple	12	Yes	1	1
432002	CoolCell LX, green	12	Yes	1	1
432003	CoolCell LX, orange	12	Yes	1	1
432004	CoolCell LX, pink	12	Yes	1	1
432138	CoolCell LX, 4 colors (purple, green, orange, pink)	12	Yes	--	4
432005	CoolCell 5 mL LX, purple	12	Yes	1	1
432006	CoolCell FTS30, purple	30	Yes	1	1
432007	CoolCell FTS30, orange	30	Yes	1	1
432008	CoolCell FTS30, green	30	Yes	1	1
432009	CoolCell FTS30, pink	30	Yes	1	1
432010	CoolCell SV2	12	Yes	1	1
432011	CoolCell SV10	6	Yes	1	1

Corning Vacuum Filtration Systems

Cat. No.	Membrane	Funnel/Bottle Volume (mL)	Pore Size (µm)	Qty/Cs
431153	PES	150/150	0.22	12
431096	PES	250/250	0.22	12
431097	PES	500/500	0.22	12
431098	PES	1,000/1,000	0.22	12

Corning Bottle Top Vacuum Filtration Systems

Cat. No.	Membrane	Volume (mL)	Neck Size (mm)	Pore Size (µm)	Qty/Cs
431161	PES	150	45	0.22	48
431118	PES	500	45	0.22	12
431174	PES	1,000	45	0.22	12

Corning Syringe Filters

Cat. No.	Diameter (mm)	Pore Size (µm)	Housing Material	Membrane Material	Sterile	Inlet/Outlet	Packaging	Qty/Cs
431219	28	0.2	SFCA	AC	Yes	LL/LS	Indiv.	49
431222	25	0.2	RC	PP	Yes	LL/LS	Indiv.	50
431224	25	0.2	NY	PP	Yes	LL/LS	Indiv.	50
431229	20	0.2	PES	AC	Yes	LL/LS	Indiv.	50

Reagents

Cat. No.	Description	Size	Qty/Cs
25-950-CQC	Dimethyl Sulfoxide	250 mL	1
354253	Cell Recovery Solution	100 mL	1
354216	Calcein AM fluorescent dye, 500 µg (10 x 50 µg)	500 mg	1
354217	Calcein AM fluorescent dye	1 mg	1
354218	DiIC12(3) fluorescent dye	100 mg	1
25-900-CI	Trypan Blue Solution, 0.4% (w/v) in PBS, pH 7.5 ± 0.5	100 mL	1
354235	Dispase	100 mL	1

Sera

Cat. No.	Description	Size	Qty/Cs
35-010-CV	Fetal Bovine Serum, regular	500 mL	1
35-015-CV	Fetal Bovine Serum, premium	500 mL	1

Corning Media

Cat. No.	Description	Size	Qty/Cs
10-013-CV	DMEM [+], 4.5 g/L glucose, L-glutamine, sodium pyruvate	500 mL	6
10-017-CV	DMEM [+], 4.5 g/L glucose, L-glutamine [-], sodium pyruvate	500 mL	6
15-013-CV	DMEM [+], 4.5 g/L glucose, sodium pyruvate [-], L-glutamine	500 mL	6
15-090-CV	DMEM/F12 50:50 mix [-], L-glutamine	500 mL	6
10-092-CV	DMEM/Ham's F-12 50/50 mix [+], L-glutamine, 15 mM HEPES	500 mL	6
16-405-CV	DMEM/Ham's F-12 50/50 mix [+], L-glutamine [-], phenol red	500 mL	6
15-016-CV	Iscove's Modification of DMEM [+], 25 mM HEPES, [-], β-thioglycerol, β-mercaptoethanol, L-glutamine	500 mL	6
10-016-CV	Iscove's Modification of DMEM [+], L-glutamine, 25 mM HEPES [-], β-thioglycerol, β-mercaptoethanol	500 mL	6
10-080-CV	Ham's F-12 Medium [+], L-glutamine	500 mL	6
10-060-CV	Medium 199 (Mod.) [+], Earle's salts, L-glutamine	500 mL	6
10-022-CV	MEM alpha medium [+], Earle's salts, ribonucleosides, deoxyribonucleosides, L-glutamine	500 mL	6
15-012-CV	MEM alpha medium [+], Earle's salts [-], ribonucleosides, deoxyribonucleosides, L-glutamine	500 mL	6
10-010-CV	MEM [+], Earle's salts, L-glutamine	500 mL	6
10-009-CV	MEM [+], 1.5 g/L sodium bicarbonate, NEAA, L-glutamine, sodium pyruvate	500 mL	6
15-010-CV	MEM [+], Earle's salts [-], L-glutamine	500 mL	6
17-305-CV	MEM [+], Earle's salts [-], L-glutamine, phenol red	500 mL	6
10-040-CV	RPMI 1640 [+], L-glutamine	500 mL	6
15-040-CV	RPMI 1640 [-], L-glutamine	500 mL	6
17-105-CV	RPMI 1640 [-], L-glutamine, phenol red	500 mL	6
10-041-CV	RPMI 1640 [+], L-glutamine, 25 mM HEPES	500 mL	6
15-041-CV	RPMI 1640 [+], 25 mM HEPES [-], L-glutamine	500 mL	6

Corning Buffers

Cat. No.	Description	Size	Qty/Cs
21-040-CV	PBS (Phosphate-Buffered Saline), 1X without calcium and magnesium	500 mL	6
21-020-CV	HBSS (Hanks' Balanced Salt Solution), 1X with calcium and magnesium	500 mL	6
21-023-CV	HBSS, 1X with calcium and magnesium without phenol red	500 mL	6
21-021-CV	HBSS, 1X without calcium and magnesium	500 mL	6
21-022-CV	HBSS, 1X without calcium, magnesium, and phenol red	500 mL	6
21-030-CV	DPBS (Dulbecco's Phosphate-Buffered Saline), 1X with calcium and magnesium	500 mL	6
21-031-CV	DPBS, 1X without calcium and magnesium	500 mL	6
25-015-CI	Corning glutagro™ supplement	100 mL	1
25-021-CI	Trace Elements A, 1,000X	100 mL	1
25-022-CI	Trace Elements B, 1,000X	100 mL	1
25-023-CI	Trace Elements C, 1,000X	100 mL	1
25-030-CI	MEM amino acids, 50X	100 mL	6
25-025-CI	MEM Nonessential amino acids, 100X	100 mL	6
25-800-CR	ITS (Insulin-Transferrin-Selenium)	10 mL	1

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